erence in their entirety, describe the use of a continuous polymer bed formed by in situ polymerization of a monomer solution containing a porogen within a column. Many examples on the use of these continuous or monolithic polymer supports are available in the literature. Liao in U.S. Pat. No. 5,647,979 describes a similar use of a continuous polymer bed for reversed-phase chromatography and capillary electrochromatography in capillary columns.

[0012] Detection of analytes separated on an LC column has traditionally been accomplished by use of spectroscopic detectors. Spectroscopic detectors rely on a change in refractive index, ultraviolet and/or visible light absorption, or fluorescence after excitation with a suitable wavelength to detect the separated components. Additionally, the effluent from an LC column may be nebulized to generate an aerosol which is sprayed into a chamber to measure the light scattering properties of the analytes eluting from the column. Alternatively, the separated components may be passed from the liquid chromatography column into other types of analytical instruments for analysis. The volume from the LC column to the detector is minimized in order to maintain the separation efficiency and analysis sensitivity. All system volume not directly resulting from the separation column is referred to as the dead volume or extra-column volume.

[0013] The miniaturization of liquid separation techniques to the nano-scale involves small column internal diameters (<100 μm i.d.) and low mobile phase flow rates (<300 nL/min). Currently, techniques such as capillary zone electrophoresis (CZE), nano-LC, open tubular liquid chromatography (OTLC), and capillary electrochromatography (CEC) offer numerous advantages over conventional scale high performance liquid chromatography (HPLC). These advantages include higher separation efficiencies, high-speed separations, analysis of low volume samples, and the coupling of 2-dimensional techniques. One challenge to using miniaturized separation techniques is detection of the small peak volumes and a limited number of detectors that can accommodate these small volumes. However, coupling of low flow rate liquid separation techniques to electrospray mass spectrometry results in a combination of techniques that are well suited as demonstrated in J. N. Alexander IV, et al., *Rapid Commun. Mass Spectrom.* 12:1187-91 (1998). The process of electrospray at flow rates on the order of nanoliters ("nL") per minute has been referred to as "nanoelectrospray".

[0014] Capillary electrophoresis is a technique that utilizes the electrophoretic nature of molecules and/or the electroosmotic flow of fluids in small capillary tubes to separate components of a fluid. Typically, a fused silica capillary of 100 μm inner diameter or less is filled with a buffer solution containing an electrolyte. Each end of the capillary is placed in a separate fluidic reservoir containing a buffer electrolyte. A potential voltage is placed in one of the buffer reservoirs and a second potential voltage is placed in the other buffer reservoir. Positively and negatively charged species will migrate in opposite directions through the capillary under the influence of the electric field established by the two potential voltages applied to the buffer reservoirs. Electroosmotic flow is defined as the fluid flow along the walls of a capillary due to the migration of charged species from the buffer solution under the influence of the applied electric field. Some molecules exist as charged species when in solution and will migrate through the

capillary based on the charge-to-mass ratio of the molecular species. This migration is defined as electrophoretic mobility. The electroosmotic flow and the electrophoretic mobility of each component of a fluid determine the overall migration for each fluidic component. The fluid flow profile resulting from electroosmotic flow is flat due to the reduction in frictional drag along the walls of the separation channel. This results in improved separation efficiency compared to liquid chromatography where the flow profile is parabolic resulting from pressure driven flow.

[0015] Capillary electrochromatography is a hybrid technique that utilizes the electrically driven flow characteristics of electrophoretic separation methods within capillary columns packed with a solid stationary phase typical of liquid chromatography. It couples the separation power of reversed-phase liquid chromatography with the high efficiencies of capillary electrophoresis. Higher efficiencies are obtainable for capillary electrochromatography separations over liquid chromatography, because the flow profile resulting from electroosmotic flow is flat due to the reduction in frictional drag along the walls of the separation channel when compared to the parabolic flow profile resulting from pressure driven flows. Furthermore, smaller particle sizes can be used in capillary electrochromatography than in liquid chromatography, because no backpressure is generated by electroosmotic flow. In contrast to electrophoresis, capillary electrochromatography is capable of separating neutral molecules due to analyte partitioning between the stationary and mobile phases of the column particles using a liquid chromatography separation mechanism.

[0016] Microchip-based separation devices have been developed for rapid analysis of large numbers of samples. Compared to other conventional separation devices, these microchip-based separation devices have higher sample throughput, reduced sample and reagent consumption, and reduced chemical waste. The liquid flow rates for microchip-based separation devices range from approximately 1-300 nanoliters per minute for most applications. Examples of microchip-based separation devices include those for capillary electrophoresis ("CE"), capillary electrochromatography ("CEC") and high-performance liquid chromatography ("HPLC") include Harrison et al., *Science* 261:859-97 (1993); Jacobson et al., *Anal. Chem.* 66:1114-18 (1994), Jacobson et al., *Anal. Chem.* 66:2369-73 (1994), Kutter et al., *Anal. Chem.* 69:5165-71 (1997) and He et al., *Anal. Chem.* 70:3790-97 (1998). Such separation devices are capable of fast analyses and provide improved precision and reliability compared to other conventional analytical instruments.

[0017] The work of He et al., *Anal. Chem.* 70:3790-97 (1998) demonstrates some of the types of structures that can be fabricated in a glass substrate. This work shows that co-located monolithic support structures (or posts) can be etched reproducibly in a glass substrate using reactive ion etching (RIE) techniques. Currently, anisotropic RIE techniques for glass substrates are limited to etching features that are 20 μm or less in depth. This work shows rectangular 5 μm by 5 μm width by 10 μm in depth posts and stated that deeper structures were difficult to achieve. The posts are also separated by 1.5 μm. The posts supports the stationary phase just as with the particles in LC and CEC columns. An advantage to the posts over conventional LC and CEC is that